Detection of *Pneumocystis jirovecii* by Grocott-Gomori Methenamine Silver and Indirect Immunofluorescence Assay in Chronic Obstructive Pulmonary Disease (COPD) Patients

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Abstract

Background: *Pneumocystis jirovecii* has emerged as an opportunistic fungal pathogen in immunocompromised patients. The organism has a tropism to colonized lung in patients with chronic pulmonary obstructive disease (COPD) even with no obvious immunosuppressive state.

Aims: This study aimed to investigate the colonization of *P. jirovecii* in patients with COPD and the association of different demographic characteristics with this colonization.

Patients and Methods: Sputum samples were collected from 100 patients with COPD. Samples were processed according to the standard protocols and were examined by two laboratory techniques: Grocott-Gomorimethenaminesilver (GMS) and indirect immunofluorescence assay (IFA). Data including age, gender, smoking status and hospital stay were extracted from patients through direct interview.

Results: The overall detection rate of *P. jirovecii* from COPD patients using GMS and IFA was 8% and 9% respectively. Older ages and smoking were associated significantly with increased risk of *P. jirovecii* colonization in COPD patients.

Conclusions: These data indicate the importance of *P. jirovecii* as opportunistic pathogen in colonization of the respiratory tract of patients with COPD. Further studies are required to illustrate the clinical implication of this colonization.

Keywords: *Pneumocystis jirovecii*, chronic obstructive pulmonary disease (COPD).

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Introductions

COPD is a progressive lung diseases associated with a chronic inflammatory response involving airways and lung parenchyma [1]. Recent reports considered COPD as the fourth leading cause of mortality worldwide. With less severe cases, there are heavy health burden and high medical cost[2]. Both genetic and environmental factors overlap to introduce COPD. Beside smoking, recent evidence supports the role of infectious agents in the triggering and perpetuating of COPD [3]. *P. jirovecii* is an atypical opportunistic organism belonging to the fungi according to the phylogenetic studies based on whole genome sequencing [4,5]. This organism has a tropism to pulmonary tissues and causes pneumonia especially in immunocompromised individuals [6]. Several evidence has linked *P. jirovecii* with COPD. These include COPD-like feature that take place after pneumocystis pneumonia (PCP), the inflammatory response in COPD and *Pneumocystis* colonization is alike, and the occurrence of airway obstruction in animal models subjected to pneumocystis colonization [3]. It seem that the colonization of *P. jirovecii* is not

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associated with impaired function of lung; rather the organism especially colonies the lung with COPD. Probst *et al.*reported 7% and 40% colonization of *P. jirovecii* in cystic fibrosis and COPD respectively [7]. Furthermore, Helweg-Larsen *et al.* found 4% of patients with suspected bacterial pneumonia were positive for *P. jirovecii*, 63% of whom had COPD, while only 20% of COPD free patients had such colonization [8].Regarding the source of infection, recent evidence suggested a continuous acquisition of the micro-organism rather than the presence of latent infection. Supporting this assumption is the frequent reported outbreaks in different settings like liver transplant units [9], pediatric oncology wards [10] and hematology ward [11] where *P. jirovecii*DNA was detected in the air surrounding patients with or without active PCP.

There is paucity in the studies regarding the colonization rate of *P. jirovecii*in COPD patients in Iraq. Therefore, this study aimed to investigate the colonization of this fungus in human immunodeficiency virus (HIV)-negative COPD patients and its association with some demographic factors.

Subjects and Methods:

The Study Population and Sampling

Sputum samples were collected from 100 moderate to severe COPDpatients who were admitted to respiratory ward in AL- Imamian AL- KadhumainMedical City during the period from May-2017 to March-2018.Inclusion criteria were all tpatients with moderate to severe COPD with sera-negative for HIV infection by ELISA and western blot test. Moderate to severe COPD was defined according to the Global Initiative for Chronic Obstructive Lung Disease (GOLD) criteria (a ratio of forced expiratory volume in 1 second [FEV1] to forced vital capacity [FVC] of <70% after bronchodilator use and an FEV1 of <80% of the predicted value after bronchodilator use). Clinical manifestations were determined by consultation of Respiratory and Chest diseases specialist. Exclusion criteria were patients diagnosed with asthma or bronchiectasis at the sampling, patients with immunosuppressive drugs, and those with life-threatening disorders. The ethical aspects of this study have been approved by the ethical council in Medical College,Al- Nahrain University.

Samples Collection and Processing:

Sputum samples containing mucous material were added to a 2-fold volume of 0.9% NaCl and mixed by vortexing for 5 minutes. The mixture was then centrifuged at 3000 rpm for 5 minutes, and the supernatants were discarded. Two laboratory methods were used for *P. jirovecii*detection in this study. The first method is staining with GMS stain (abcam®, UK). Aportion of precipitate pellets (100µl) was used to prepare smears which were fixed with ethanol and stained with GMS stain according to the standard protocol[12]. The second method was the fluorescent monoclonal antibody test using a commercially ready kit (Axis-Shield Diagnostics, and Dundee, UK) according to the manufacturer's instructions. In this method, the pellets were resuspended with sterile distilled water and washed twice by centrifugation. Several slides were prepared on which smears from the resuspended pellets were prepared and fixed with high quality acetone. After drying at room temperature, the slides were rinsed with distilled water to remove the salts form the specimens. The specimens were then treated with enzyme solution containing trypsin. Monoclonal murine anti-*P. jirovecii* antibody and fluorescently labelled anti-mouse antibody were added and the preparation was incubated at 37°C for 10 min. in a humidified chamber. After incubation, the slides were rinsed and air-dried and examined at ×100 magnification using fluorescent microscope. A positive finding was based on the detection of the fluorescent cysts which have round or elliptical shape[13].

Statistical Analysis

Continuous variables were expressed as mean± standard deviation (SD) and analyzed with student t-test, while binomial variables were expressed as frequency and percentages, and analyzed with Chi-square. All analyses were conducted with SPSS software. A p-value of 0.05 or less was considered significant.

Results

Demographic and Clinical Characteristics of Patients

The demographic and clinical characteristic of COPD patients are shown in table 1. Most patients were elderly, and relatively more females (57%) were affected than males (43%). This high percentage of female was reflected by lower percentage of smokers (38%) than non-smokers (62%) because most women re non-smokers. Most patients

have 3 days or less hospital stayingCough was predominant and reported in 83% of patients, while about two-third of patients were suffering from shortness of breath.

Table 1: Demographic and clinical characteristics of the study population

Variables	Value, Frequency(%)	
Age, year (mean±SD)	66.5 ± 11	
Gender		
Male	43(43%)	
Female	57(57%)	
Hospital stay		
≤3 days	72(72%)	
4 days or more	28(28%)	
Smoking		
No	62(62%)	
Yes	38(38%)	
Cough	83(83%)	
Shortness of breath	69(69%)	
Fever	26(26%)	

Detection of Pneumocystis jiroveci

Grocott-GomoriMethenamine Silver Stain

Cysts of *P. jiroveci* is the diagnostic form, and under light microscope, it had a characteristic appearance as a spherical, or cup-shapedobjects when stained by GMS (Figure 1). Out of 100sputum samples of COPD patients, the organism was detected in 8(8%) with this technique.

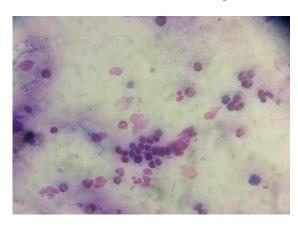


Figure 1: P. jirovecii cysts detected by GMS stain (×100).

Immunofluorescence Detection of Pneumocystis jirovecii

Under fluorescent microscope, *P. jirovecii* cysts appear as round or elliptical structure with green color (Figure 2). Out of 100sputum samples of COPD patients, the organism was detected in 9(9%) samples by IFA test. Of note, all specimens gave positive results for GMS Stain were positive for fluorescent test, which implies the superiority of IFA monoclonal antibody over GMS stain in detection of *P. jirovecii* cyst.

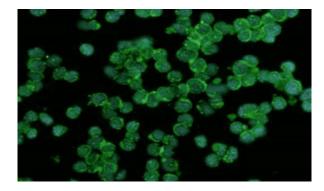


Figure (2): Immunofluorescent staining using monoclonal antibodies against *P. jirovecii*. the cyst appears bright to bright apple green($\times 100$).

Effect of Demographic Characteristics on P. jirovecii Colonization

The association of different demographic characteristic with the colonization of P. jiroveciiwas further investigated (Table 2). Both age and smoking status were found to be significantly influence such colonization. Mean age of infected group was 82.44 ± 18.2 years versus 59.92 ± 9.3 years (P=0.016). More than three-quarters of infected patients were smokers compared to 34.07% among non-infected (P=0.011).

Table 2: Association of P. jirovecii infection with some demographic characteristics

Variables	`Infected	Non-infected	
	(9)	(91)	P-value
Age, year (mean±SD)	82.44 ± 18.2	59.92±9.3	0.016
Gender			
Male	5(55.56%)	38(43.95%)	0.428
Female	4(44.44%)	53(56.05%)	
Hospital stay			
≤3 days	4(44.44%)	68(74.73%)	0.068
4 days or more	5(55.56%)	23(25.27%)	
Smoking			
No	2(22.22%)	60(65.93%)	0.011
Yes	7(77.78%)	31(34.07%)	

Discussion

Pneumocystis jirovecii is an opportunistic pathogen which colonizes lung and associates with COPD. Detection of this colonization is very important for two main reasons. Firstly for better understanding of the organism epidemiology and its correlation with lung disease. Secondly, most cases of colonization represent starting points for PCP especially when there is an immune suppression [14]. The current study reported 9% incidence P. jirovecii in patients with COPD. These results are completely in accordance with that obtained in a local study conducted Yehiaet al. on patients with lower respiratory tract infections [15]. In contrast, Touhali et al. reported14% of immunocompromised patients were positive for Pneumocystosis [16]. This high rate is reasonable because the immune status of the study population.Of note, in these two studies, conventional (not molecular) methods were used for detection of the fungus. Globally, Sheikholeslami et al. in Iran reported very close rate (7%) of infection in patients withwith COPD [17]. More recently, in Iran also, Aboualigalehdari et al. subjected 115 BAL specimens from patients with different chronic pulmonary disorders for nested-polymerase chain reaction with specific primers for P. jirovecii. They found that 27% of their patients were positive for this opportunistic pathogen [18]. It is obvious form these studies and others that there are several factors which influence the detection rate. These include age, complain and immune status of the patients, specimens type, and, may be the most important, the method of

detection. In this regard, Ng et al. reported that samples were considered positive to P. jirovecii (i.e., a truly positive specimen) if this organism was detected by two or more of the staining methods [19]. Indeed, this was true before the era of PCR which dramatically changed the standard of diagnosis. In a recent study, Moodly et al. compared the efficiency of quantitative PCR (qPCR) with IFA in detection of P. jirovecii in sputum samples from patients suspected to have PCP. As expected, qPCR had higher detection rate than IFA (67% vs 51%) [20]. However, this preponderance of molecular methods is not predominant. In an Egyptian study including 50 children with hematological malignancies, three diagnostic methods (Giemsa staining, PCR and direct IFA) were used for P. *jirovecii* detection. Interestingly, IFA was found to have superiority over PCR or Giemsa staining in this study [21]. In fact, even within molecular methods there is some variation in detection rate, and there is a general agreement that real-time PCR is more sensitive than conventional PCR. The current study revealed a significant positive association of age and smoking with the colonization of P. jirovecii. For age association, a previous Japanese report confirmed such influence [22]. Another study involving 110 elderly HIV-negative subjects attending a preventive health care program detected P. jirovecii in 12.8% which is a relatively high rate [23]. Two factors have been proposed to explain the high incidence of *P. jirovecii* in elderly, both of them related to the immune status. These are the decline in cellular immune response accompanying the aging process (known as immunosenescence), and the medications taken by elderly which may have negative impact on the immune system [22, 24]. Although it was not significant in the current study, hospital stay appears to have positive impact on P. jirovecii colonization in the lung. A prolonged hospital stay increases the risk of getting nosocomial infections. In one study, it was found that healthcare workers are able to carry and transmit this pathogen for up to 10 weeks [25]. Smoking is a well-known risk factor for many respiratory infections. The current study showed a significant association between smoking and P. jirovecii colonization. This result agrees with the recent study by Tokemoto et al. on healthy smokers and nonsmokers. The authors reported a significant impact of smoking on the colonization rate of *P. jirovecii* (20% vs 47%) [26]. The exact mechanism by which smoking has such influence is no exactly clarified. However, smoking induces a mechanical damage accompanied with an increase of inflammatory cytokines from alveolar tissues [27]. These changes and may be others could create a favorable microenvironment for *P. jirovecii*colonization.

Collectively, these data indicates the significant of *P. jirovecii* in COPD patients. A cohort study investigating the effect of this colonization on lung function and the possible development to PCP is required.

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Conflict of interest

The authors declare that they have no competing interests.

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References

- 1- Khodavaisy S, Mortaz E, Mohammadi F, Aliyali M, Fakhim H, Badali H. *Pneumocystis jirovecii* colonization in chronic obstructive pulmonary disease (COPD). Curr Med Mycol 2015;1(1):42-48.
- 2- May SM, Li JTC. Burden of chronic obstructive pulmonary disease: healthcare costs and beyond. Allergy Asthma Proc 2015;36(1):4-10.
- 3- Morris A, Sciurba FC, Norris KA. Pneumocystis: a novel pathogen in chronic obstructive pulmonary disease. COPD 2008;5(1):43-51.
- 4- Cisse OH, Pagni M, Hauser PM: De novo assembly of the *Pneumocystis jirovecii* genome from a single bronchoalveolar lavage fluid specimen from a patient. MBio. 2012; 4(1): e00428–12.
- 5- Ma L, Chen Z, Huang da W, et al.: Genome analysis of three *Pneumocystis* species reveals adaptation mechanisms to life exclusively in mammalian hosts. Nat Commun. 2016; 7: 10740.
- 6- Gigliotti F, Limper AH, Wright T. Pneumocystis. Cold Spring HarbPerspect Med 2014;2(12):a019828.
- 7- Probst M, Ries H, Schmidt-Wieland T, Serr A. Detection of *Pneumocystis carinii* DNA in patients with chronic lung diseases. Eur J ClinMicrobiol Infect Dis. 2000;19:644–645.
- 8- Helweg-Larsen J, Jensen JS, Dohn B, Benfield TL, Lundgren B. Detection of *Pneumocystis* DNA in samples from patients suspected of bacterial pneumonia—a case-control study. BMC Infect Dis. 2002;2:28.

- 9- Desoubeaux G, Dominique M, Morio F, et al.: Epidemiological Outbreaks of *Pneumocystis jirovecii* Pneumonia Are Not Limited to Kidney Transplant Recipients: Genotyping Confirms Common Source of Transmission in a Liver Transplantation Unit. J ClinMicrobiol. 2016; 54(5): 1314–20.
- 10- Yiannakis EP, Boswell TC: Systematic review of outbreaks of *Pneumocystis jirovecii* pneumonia: evidence that *P. jirovecii* is a transmissible organism and the implications for healthcare infection control. J Hosp Infect. 2016; 93(1): 1–8.
- 11- Robin C, Alanio A, Gits-Muselli M, et al.: Molecular Demonstration of a *Pneumocystis* Outbreak in Stem Cell Transplant Patients: Evidence for Transmission in the Daycare Center. Front Microbiol. 2017; 8: 700.
- 12- Swisher BL, Chandler FW. Grocott-Gomorimehenamine silver method for detecting fungi: practical consideration. Lab Med 1982;13(9):568-570.
- 13- Harris JR, Marston BJ, Sangrujee N, DuPlessis D, Park B. Cost-effectiveness analysis of diagnostic options for pneumocystis pneumonia (PCP). PLoS ONE. 2011;15;6(8):e23158.
- 14- Alanio A and Bretagne S. *Pneumocystis jirovecii* detection in asymptomatic patients: what does its natural history tell us? F1000Research 2017, 6:739
- 15- Yehia MM, Al-Habbo DJ, Abdulla ZA. Detection of *Pneumocystis carinii* (*jirovecii*) from Iraqi patients with lower respiratory tract infections. Iraqi J Med Sci. 2014; 126-130.
- 16- Touhali IS, Ibrahim AA, Dawood HN. Conventional methods for the diagnosis of *Pneumocystis jirovecii* in immunocompromised Iraqi patients. Iraqi J Med Sci 2016;1:14.
- 17- Sheikholeslami MF, Sadraei J, Farnia P, Forozandeh M. Colonization of *Pneumocystis jirovecii* in chronic obstructive pulmonary disease (COPD) patients and the rate of Pneumocystis pneumonia in Iranian non-HIV+ immunocompromised patients. Iranian J Microbiol 2013;5(4):411.
- 18- Aboualigalehdari E, Mahmoudabadi AZ, Fatahinia M, Idani E. The prevalence of *Pneumocystis jirovecii* among patients with different chronic pulmonary disorders in Ahva, Iran. Iran J Microbiol 2015;7(6):333-337.
- 19- Ng VL, Yajko DM, McPhaul LW, *et al.* Evaluation of an indirect fluorescent-antibody stain for detection of Pneumocystis carinii in respiratory specimens. J ClinMicrobiol 1990;28:975–979.
- 20- Moodley B, Tempia S, Frean JA. Comparison of quantitative real-time PCR and direct immunofluorescence for the detection of *Pneumocystis jirovecii*. PLoS ONE 2017;12(7): e0180589.
- 21- El-Seidi EA, El-Hodaky SK, Ouda NH, et al. Comparison of different methods to diagnose *Pneumocystis jirovecii* pneumonia in children with hematological malignancies. Egyptian J Med Microbiol 2008;17(1):1151-159.
- 22- Mori S, Cho I, Ichiyasu H, Sugimoto M. Asymptomatic carriage of Pneumocystis jirovecii in elderly patients with rheumatoid arthritis in Japan: a possible association between colonization and development of pneumocystis jirovercii pneumonia during low-dose MTX therapy. Mod Rheumatol 2008;18(3):240-246.
- 23- Vargas S, Piarro P, Lope-Vieyra M, Neira-Aviles P, Bustamante R, Ponce CA. *Pneumocystis jirovecii* colonization in older adults and diagnostic yield of single versus paired noninvasive respiratory sampling. ClinInfec Dis 2010;50(3):e19-e21.
- 24- Effros RB, Fletcher CV, Gebo K, *et al.* Workshop on HIV infection and aging: what is a known and future research direction. Clin Infect Dis 2008; 47:542–553.
- 25- Durand-Joly I, Soula F, Chabé M, et al.: Long-term colonization with *Pneumocystis jirovecii* in hospital staffs: a challenge to prevent nosocomial pneumocystosis. J EukaryotMicrobiol. 2003; 50(Suppl): 614–5.
- 26- Takemoto S, Ebara M, Hasebe E, Yakushijin Y. A study on the colonization of *Pneumocystis jirovecii* among outpatients during cancer chemotherapy and among healthy smokers. J InfecChemother 2017 23(11):752-756.
- 27- Marshall MM, McCormack MC, Kirk GD. Effect of cigarette smoking on HIV acquisition, progression, and mortality. AIDS EducPrev 2009; 21:28e39.